

## RNA (Northern) Blot Analysis

### Procedure:

- (1) Wet membrane in 2X SSC
- (2) Pre-pre hyb 60°C    ~1h
- (3) Pre-hyb 42°C    4h - O/N
- (4) Hyb 42°C    O/N
- (5) Low stringency washes (3) RT    ~15min each
- (6) High stringency washes (3 or 4) 55°C    ~60min each
- (7) Expose film to filter
- (8) Strip membrane (2 washes) 10-15min each  
(Boil stripping solution then add to filter)

### Hybridization Solution Preparation:

	<u>Pre-pre</u> (20ml)	<u>Pre-hyb</u> (10ml)	<u>Hyb</u> (5ml)
(1) Formamide	-	5.0ml	2.5ml
(2) 25X SSC	0.8ml	2.0ml	1.0ml
(3) 2M PB	-	0.25ml	0.05ml
(4) Denhardt's	-	1.0ml	0.1ml
(5) 10% SDS	1.0ml	0.2ml	0.05ml
(6) Dextran Sulfate	-	-	1.0ml
(7) 10mg/ml Fish DNA	-	0.2ml	0.05ml
(8) dH2O	38.2ml	1.35ml	-
(9) Probe	-	-	0.5-1.0ml

### Stock Solution Preparation:

(1) **Formamide:** stock solution is deionized by stirring with AG 501-X8 (D) ion exchange resin (BIORAD) for 30 min and subsequently filtering/decanting.

#### (2) 25X SSC

	<u>Stock</u>	<u>Conc</u>
1.	NaCl <span style="float: right;">43.8g</span>	3.75M
2.	Na-Citrate <span style="float: right;">22.1g</span>	0.375M
3.	pH w/HCl to <span style="float: right;">7.0</span>	
4.	ddH <sub>2</sub> O to <span style="float: right;">250ml</span>	
5.	Filter through a 0.45μM filter, Autoclave	

#### (3) 2M PB

1. Na<sub>2</sub>HPO<sub>4</sub> 28.3g
2. ddH<sub>2</sub>O 160ml
3. Mix until dissolved (heat ok)
4. NaH<sub>2</sub>PO<sub>4</sub> 27.5g
5. Mix until thoroughly dissolved
6. Bring vol to 200ml

7. Filter through a 0.45 $\mu$ M filter, Autoclave

**(4) 100X Denhardt's**

1. BSA 0.5g
2. PVP 0.5g
3. Ficoll 0.5g
4. ddH<sub>2</sub>O to 25.0ml
5. Filter through cinder-glass using Millipore apparatus
6. Store at -20°C

**(5) 10% SDS**

1. SDS 10g
2. ddH<sub>2</sub>O to 100ml
3. Sterile filter

**(6) 50X Dextran sulfate**

1. Dextran sulfate 25g
2. Sterile H<sub>2</sub>O to 50ml
3. Heat/Vortex to dissolve
4. Store -20°C

**(7) 10 mg/ml Fish DNA**

1. Dissolve 0.1g of Fish DNA in 10ml sterile ddH<sub>2</sub>O. Shear by drawing the DNA up into a syringe and passing it out through a 22 g needle. Repeat ~ 20x.
2. Store at -20°C.
3. Boil for 10 minutes before using.

**(8) 5% PPI**

1. Sodium pyrophosphate 7.5g
2. ddH<sub>2</sub>O to 150ml
3. Filter through a 0.45 $\mu$ M filter, Autoclave

**Wash Buffers:**

<u>Stock</u>	<u>Low stringency</u>		<u>High stringency</u>	
	<u>Vol</u>	<u>Conc</u>	<u>Vol</u>	<u>Conc</u>
(1) 20X SSC	25ml	[2X]	2.5ml	[0.2X]
(2) 10% SDS	1.25ml	[0.05%]	1.25ml	[0.05%]
(3) 5% PP <sub>i</sub>	1.0ml	[0.02%]	0.5ml	[0.01%]
(4) dH <sub>2</sub> O	222.75ml		245.75ml	

**Strip Buffer:**

(1) dH <sub>2</sub> O	500ml
(2) BOIL	
(3) 10% SDS	5ml

**Note:** the above Hyb volumes are for a 10 X 15 cm filter.