

RNA Gels

(Adapted from the Red Book i.e., Ausubel)

Gel (One 1.2% Agarose, 10 X 15cm, 165ml gel):

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|------------------------|----------|
| (1) Agarose | 2g |
| (2) ddH ₂ O | 143.55ml |
| (3) Boil | |
| (4) Cool | to 60°C |
| (5) 10X MOPS | 16.5ml |
| (6) Formaldehyde | 4.95ml |

Sample Prep:

Per Sample (adjust volumes according to well volume)

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|------------------------------|--------|
| (1) 10X MOPS | 5μL |
| (2) 37% (stock) Formaldehyde | 8.75μL |
| (3) Formamide (Deionized) | 25μL |
| (4) 10mg/ml Ethidium Bromide | 0.05μL |

(5) Mix:

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|------------------------------------|---------|
| (a) RNA (2-10μg*) | ___μL |
| (b) Sample Prep Sol'n | 38.75μL |
| (c) DEPC treated dH ₂ O | to 50μL |

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|---------------------------------|-------|
| (6) Incubate @ 55-68°C | 15min |
| (7) Formaldehyde loading buffer | 10μl |

(8) Load samples onto the gel.

(9) Run ~100V ~2-3h (200 -300 V-h).

(10) Photograph the gel

(11) Rinse the gel by incubating it in ddH₂O for 30min with gentle shaking.

(12) Discard the water.

(13) Repeat the wash step with a second 30 min incubation.

(14) Set up a transfer blot. See the procedure "Genomic(Plant)DNA Restriction Digestion For Southernns" on how to set up a blot. Note that RNA is already denatured and so there is no need to run through the denaturation/neutralization steps as with Southernns. Simply follow the steps for *Transfer Membrane, Gel Blot Paper* and *Assembling the Blot*, and blot gel onto Hybond N nylon membrane overnight using 10X SSC.

(15) Disassemble the gel blot and remove the filter from the gel.

(16) Invert the filter so the DNA side is facing up and lay it on a piece of plastic wrap.

(17) Cross-link the RNA to the filter using the UV Stratalinker 2400:

- Turn on the UV Stratalinker 2400
- Place filter, DNA side up, into the chamber
- Close the door
- Press Auto Cross Link
- Press Start

Solutions and Stuff:

	<u>Stock</u>	<u>Final concentration</u>
(1) Formaldehyde loading buffer		
(a) 0.5M EDTA	2 μ L	1mM
(b) Bromophenol Blue	Dab	~0.25%
(c) Xylene Cyanol	Dab	~0.25%
(d) 80% Glycerol	625 μ L	50%
(e) dH ₂ O	373 μ L	

(2) 10X MOPS Buffer

(a) MOPS	41.8g
(b) 3M DEPC treated NaOAc	16.6ml
(c) 0.5M EDTA	20.0ml
(d) ddH ₂ O	to 1L
(e) DEPC	1ml
(f) Filter sterilize	
(g) Store at RT in a brown bottle	

Give the gel box, casting tray and comb a good wash with detergent before pouring the gel.

*We have found that a range of total RNA concentrations work for hybridization analyses. Typically we load 8 μ g barley, 5 μ g *Medicago spp.*, 8 μ g *Solanaceous spp.*, and 8-10 for *Arabidopsis thaliana*.