

***In Vitro* Mutagenesis**

I. General Procedure

1. Turn on Robocycler to warm it up.
2. Thaw all necessary components (on ice)
3. Dilute template DNA to 5ng/ μ l (5 μ g/ml).
4. Label PCR tubes.
5. Aliquot complimentary primers to respective PCR tubes. Place on ice.
6. Prepare necessary volume of reaction cocktail. Do this on ice.
7. Add reaction cocktail to tubes with complimentary primers.
8. Place in Robocycler and Start PCR process.

II. Reaction Conditions:

	<u>Vol to use</u>	<u>Final Rxn []</u>	<u>Rxn Cocktail (for 5 rxns)</u>
DNA template	5.0 μ l	(25ng)	25.0 μ l
10X Reaction Buffer	5.0 μ l	(1X)	25.0 μ l
dNTP mix	1.0 μ l		5.0 μ l
DMSO	2.5 μ l	5%	12.5 μ l
ddH ₂ O (to final vol of 50 μ l)	33.0		165.0
<i>PfuTurbo</i> Polymerase	1.0 μ l	2.5U	5.0 μ l
Oligo #1	1.25 μ l	125ng	-
Oligo #2	1.25 μ l	125ng	-

(No need to overlay sample with mineral oil when using the Robocycler fitted with the Hot Top)
(Primers are assumed to be at 100ng/ μ l)

III. PCR Conditions (for a 4.5kb plasmid):

<u>Segment</u>	<u>Cycles</u>	<u>Temperature</u>	<u>Duration</u>
1	1	95°C	60 sec
2	18	95°C	30 sec
		55°C	60 sec
		68°C	9.0 min (2min/kb plasmid)
3	1	6°C	∞

IV. *DpnI* Digestions:

PCR Product	50 μ l
<i>DpnI</i> enzyme	1 μ l
37°C	~1h

Remove approximately 10% of the digest reaction to a second microfuge tube
Add 2 μ l loading dye
Run on gel
Transform 5 μ l of PCR product into 50 μ l of CaCl₂ competent *E. coli*DH5 α
Store remainder of PCR reaction product at -20°C.